

Francesco Russo

CNR Naples

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Outline

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• DNA, Sequencing, Sanger Sequencing, Next Generation Sequencing.

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- Gene, Transcription, RNA, Gene Expression, Isoform Expression,

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- Gene, Transcription, RNA, Gene Expression, Isoform Expression,
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- Alignment, Gene Expression Quantification, Coverage,
- Computational Challenges, Questions of Interest,
- Some Open Problems.

What is DNA?

DNA (Deoxy Ribonucleic Acid) is a informational molecule made from repeating units called *nucleotides*: Adenine, Guanine, Cytosine, Thymine.



What is Sequencing?

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Sequencing is the process of determining the precise order of nucleotides within a DNA molecule.

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It can be used to determine the sequence of individual genes, chromosomes or entire genomes.

Sequencing process



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Because of DNA's negative charge, the fragments move across the gel toward the positive electrode. The shorter the fragment, the faster it moves.

Sanger sequencing

Shortest fragments make the furthest progress. By considering the results of our race, we can reconstruct the nucleotide sequence.



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Sanger sequencing

Each tube contains different length fragments.



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The output Sanger sequencing machines are FASTA files

FASTA format is a text-based format for representing nucleotide sequences, in which nucleotides are represented using single-letter codes.

gi|9626685|ref|NC 001477.1| Dengue virus type 1, complete genome TTTTTAGAGAGCAGATCTCTGATGAACAACCAACGGAAAAAAGACGGGTCGACCGTCTTTCAATATGC TGAAACGCGCGAGAAACCGCGTGTCAACTGTTTCACAGTTGGCGAAGAGATTCTCAAAAGGATTGCTTTC AGOOD AGGACCONTON ANTEGET ON TOGOTTET AT AGONTTOOT A AGATTET AGOO AT ACCEDED ACA GCAGGAATTTTGGCTAGATGGGGCTCATTCAAGAAGAATGGAGCGATCAAAGTGTTACGGGGTTTCAAGA AAGAAATCTCAAACATGTTGAACATAATGAACAGGAGGAAAAGATCTGTGACCATGCTCCTCATGCTGCT AGAGGAAAATCACTTTTGTTTAAGACCTCTGCAGGTGTCAACATGTGCACCCTTATTGCAATGGATTTGG GAGAGTTATGTGAGGACACAATGACCTACAAATGCCCCCCGGATCACTGAGACGGAACCAGATGACGTTGA CTGTTGGTGCAATGCCACGGAGACATGGGTGACCTATGGAACATGTTCTCAAACTGGTGAACACCGACGA GACAAACGTTCCGTCGCACTGGCACCACGTAGGGCTTGGTCTAGAAACAAGAACCGAAACGTGGATGT CCTCTGAAGGCGCTTGGAAACAAATACAAAAAGTGGAGACCTGGGCTCTGAGACACCCAGGATTCACGGT ATGCTGGTAACTCCATCCATGGCCATGCGGTGCGTGGGAATAGGCAACAGAGACTTCGTGGAAGGACTGT CAGGAGCTACGTGGGTGGATGTGGTACTGGAGCATGGAAGTTGCGTCACTACCATGGCAAAAGACAAACC AACACTGGACATTGAACTCTTGAAGACGGAGGTCACAAACCCTGCCGTCCTGCGCAAACTGTGCATTGAA GCTAAAATATCAAACACCACCACCGATTCGAGATGTCCAACAAGGAGAAGCCACGCTGGTGGAAGAAC AGGACACGAACTTTGTGTGTCGACGAACGTTCGTGGACAGAGGCTGGGGCAATGGTTGTGGGCTATTCGG GAAAACTTAAAATATTCAGTGATAGTCACCGTACACACTGGAGACCAGCACCAAGTTGGAAATGAGACCA CAGAACATGGAACAACTGCAACCATAACACCTCAAGCTCCCACGTCGGAAATACAGCTGACAGACTACGG AGCTCTAACATTGGATTGTTCACCTAGAACAGGGCTAGACTTTAATGAGATGGTGTTGTTGACAATGAAA

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FASTA file are easy to manipulate by using scripting languages such as: Python, AWK, and Perl.

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- Main problem: most genomes are enormous (e.g 3 · 10⁹ base pairs in case of human genome, for each strand of DNA). Therefore, it is impossible for DNA to be sequenced directly.
- This is called Large-Scale Sequencing.
- A very good video about Sanger sequencing can be found here:

http://www.youtube.com/watch?v=bEFLBf5WEtc

What is Next Generation Sequencing?

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Cost and time are decreasing at each new release, while speed, accuracy and resolution are improving dramatically.

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- An Introduction to Next-Generation Sequencing Technology, from illumina.com

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NGS sequencing methodologies are able to generate hundreds of millions of short *reads* (e.g. 30 bps - 100 bps) with their quality values.
Data produced after a typical NGS process

FASTQ format is a text-based format for storing both a biological sequence and its corresponding quality scores.

```
@HWI-1KL111:53:D176AACXX:1:1101:1221:1861 1:N:0:CGATGT
NTGCTTCCTCAAAAAATTACAAAAATGCCAGTGGAGTTGTGAACTCTTCACCTCGAAGTCATAGCGCCACAAATG
#4=DDFFFHHHGHJIIIJJIJJJJJJHHIJJJFHIHGDGHIJJIJJGJJJJJIJI<FHHHIGGIJHHFFFEDEEC
@HWI-1KL111:53:D176AACXX:1:1101:1197:1873 1:N:0:CGATGT
NTGATGCTTTCAACTGCATACTTAAAGCTTGCTCCAGTTTTGTCCATCTTGTTTAAAAAACAGATTCGAGGAAAA
@HWI-1KL111:53:D176AACXX:1:1101:1228:1956 1:N:0:CGATGT
#1=DDFDDHHHHHJJJJJJJIIJJJIJIGHHIGE@GHGHIIIIIJJJIIIFHEGIE<EDFFFFFFECEEECDD=CC
@HWI-1KL111:53:D176AACXX:1:1101:1371:1853 1:N:0:CGATGT
#1=DDFFFHDFHHIGIIJIGIGHJHHHGB<>B3=BDDDD0<?B@BBBBD8B5<8B8BBBDDBB<CCACCADD:?>
@HWI-1KL111:53:D176AACXX:1:1101:1492:1889 1:N:0:CGATGT
NTATTTTTAGTAGAGACTGGATTTCTCCATGTTGGTCAGACAGGTCTCGAACTCCTGACCTCAGGTGATCTGCCT
#1=DDFFFFHHHFIIJIJJJGIJJJJJDBIIIEHIBGIIJJJIJBGIIJIDGGIJIJJIHJIEII7@GIIJJJIC
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However...

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 \ldots as the quantities of sequence data increase exponentially thanks to NGS methodologies, \ldots

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... the analysis of such a large data (usually hundreds of GigaBytes) became the major "bottom-neck" of experimental investigations.

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Pre-mRNA first has to undergo a series of modifications to become a mature *mRNA*. Differently from DNA, RNA is very perishable.

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Splicing is a modification in which introns are removed and exons are joined.

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UTR stands for Untranslated Region.

Protein production



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Protein production



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Image: A matrix

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Alternative Splicing

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 Therefore, Alternative Splicing extends the complexity of gene expression.

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These products are often proteins, but they might also be functional RNA. Therefore, Alternative Splicing extends the amount of protein production. A way to display and navigate across annotated genes is using a free software called: Genome Browser.



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The Gene Transfer Format is a file format used to hold information about gene structure.

chr1	protein_coding	exon	34554	35174	- C	() e (gene_id	"ENSG00000237613"; transcript_id "ENST00000417324";
chr1	protein coding	stop_codon		35138	35140			0	gene id "ENSG00000237613"; transcript id "ENST00000417324";
chr1	protein_coding	CDS	35141	35174		-	1	gene_id	"ENSG00000237613"; transcript_id "ENST00000417324";
chr1	processed trans	cript	exon	35245	35481		100		gene id "ENSG00000237613"; transcript id "ENST00000461467";
chr1	protein coding	CDS	35277	35481		-	2	gene id	"ENSG00000237613"; transcript id "ENST00000417324";
chr1	protein coding	exon	35277	35481		12		gene id	"ENSG00000237613"; transcript id "ENST00000417324";
chr1	processed transcript exon			35721	36073		-		gene id "ENSG00000237613"; transcript id "ENST00000461467";
chr1	protein coding	CDS	35721	35736		-	0	gene id	"ENSG00000237613"; transcript id "ENST00000417324";
chr1	protein_coding	exon	35721	36081		-		gene id	"ENSG00000237613"; transcript_id "ENST00000417324";
chr1	protein_coding	start_d	codon	35734	35736			0	gene_id "ENSG00000237613"; transcript_id "ENST00000417324";
chr1	protein_coding	CDS	69091	70005		+	0	gene_id	"ENSG00000186092"; transcript_id "ENST00000335137";
chr1	protein_coding	exon	69091	70008		+		gene_id	"ENSG00000186092"; transcript_id "ENST00000335137";
chr1	protein_coding	start_	codon	69091	69093		+	0	gene_id "ENSG00000186092"; transcript_id "ENST00000335137";
chr1	protein_coding	n_coding stop_codon			70008		+	0	gene_id "ENSG00000186092"; transcript_id "ENST00000335137";
chr1	protein_coding	exon	367640	368634		+		gene_id	"ENSG00000235249"; transcript_id "ENST00000426406";
chr1	protein_coding	CDS	367659	368594		+	0	gene_id	"ENSG00000235249"; transcript_id "ENST00000426406";
chr1	protein coding	start_codon		367659	367661		+	0	gene id "ENSG00000235249"; transcript id "ENST00000426406";
chr1	protein coding	stop_codon		368595	368597		+	0	gene_id "ENSG00000235249"; transcript_id "ENST00000426406";
chr1	protein_coding	exon	621059	622053		-		gene_id	"ENSG00000185097"; transcript_id "ENST00000332831";
chr1	protein_coding	stop_codon		621096	621098			0	gene_id "ENSG00000185097"; transcript_id "ENST00000332831";
chr1	protein_coding	CDS	621099	622034		-	0	gene_id	"ENSG00000185097"; transcript_id "ENST00000332831";
chr1	protein_coding	start_codon		622032	622034			0	gene_id "ENSG00000185097"; transcript_id "ENST00000332831";
chr1	protein_coding	exon	860260	860328		+		gene_id	"ENSG00000187634"; transcript_id "ENST00000420190";
chr1	protein_coding	exon	860530	860569		+		gene_id	"ENSG00000187634"; transcript_id "ENST00000437963";
chr1	protein_coding	exon	861118	861180	1	+		gene_id	"ENSG00000187634"; transcript_id "ENST00000342066";
chr1	protein coding	exon	861302	861393		+		gene id	"ENSG00000187634"; transcript id "ENST00000342066";
chr1	protein coding	exon	861302	861393		+		gene id	"ENSG00000187634"; transcript id "ENST00000420190";
chr1	protein coding	exon	861302	861393		+		gene id	"ENSG00000187634"; transcript id "ENST00000437963";
chr1	protein_coding	CDS	861322	861393		+	0	gene_id	"ENSG00000187634"; transcript_id "ENST00000342066";
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chr1	protein_coding	CDS	861322	861393		+	0	gene_id	"ENSG00000187634"; transcript_id "ENST00000437963";

RNA-SEQ refers to the technologies used to sequence *cDNA* in order to get information about the corresponding RNA.

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Introns are not sequenced.

Alignment

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- Alignment is a useful task to study gene expression.
- RNA alignment, also called Transcript alignment, is particularly challenging due to splicing.



Given that the transcriptome is built from the genome, the most commonly used approach is to use the genome itself as the reference.

• Uniquely mapped reads,

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- Multiple mapped reads,

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- Multiple mapped reads,
- Unmapped reads.

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Aligner tools can also make use of paired-end reads.

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Aligner tools can also make use of paired-end reads.

A *Paired-end read* is a read that has been sequenced from each end of the fragment (usually called R1 and R2).

Paired-end reads are used to increase the mapping accuracy and to reduce the number of multiple mapped reads.

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- Many different *biological variations* might be present such as: SNPs (Single-Nucleotide Polymorphism), indels, structural variations and rearrangements.

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- Many different *biological variations* might be present such as: SNPs (Single-Nucleotide Polymorphism), indels, structural variations and rearrangements.
- Reads can be affected by sequencing errors.
- Therefore, the real task is to find the location where each short read best matches the reference, while allowing for errors and structural variations.

Alignment output: SAM format

HWI-1KL111:53:D176AACXX:1:1314:7404:89056 89 chr1 10535 3 75M * 0 0 GTACCACCGAAATCTGTGCAGAGGAGGAGCACGCAGCTCCGCCCTCGCGGTGCTCCCGGGTCTGTGCTGAGGAGAAC <7BBBBB=>@EFBIIFBFECCFGD?AEFIFIIFEIIFFDFFBDA=4+1= AS:i:-5 XN:i:0 XM:i:1 X0:1:0 XG:1:0 NM:1:1 YT:7:UU NH:i:2 (C:7:chr15 (P:i:102520475 HT:i:0 MD:7:25C49 HWI-1KL111:53:D176AACXX:1:2105:1895:21247 11718 419 chr1 11650 3 75M = 143 TGGATTTTTGCCAGTCTAACAGGTGAAGCCCTGGAGATTCTTATTAGTGATTTGGGCTGGGGCCTGGCCATGTNN ?=?BDDDAF@<DFGFDBGIHIJGEFH<<FE F3:CGIA9??4?9B*?EGHGIGHIBA@@E::EECA@?### AS:i:-2 XN:i:0 XM:i:2 XO:i:0 XG:i:0 NM:i:2 MD:7:73G0T0 YT:Z:UU NH:i:2 CC:Z:chr15 CP:i:102519446 HI:i:0 HWT-1KL111:53:D176AACXX:1:2210:10686:50410 75M 329 chr1 11672 0 GTGAAGCCCTGGAGATTCTTATTAGTGATTTGGGCTGGGGCCTGGCCATGTGTATTTTTTAAATTTCCACTGAT AS:i:0 XN:i:0 XM:1:0 X0·i·0 XG:i:0 NM:i:0 MD:Z:75 YT:Z:UU NH:i:3 CC:Z:chr15 CP:i:102519424 HI:i:0 HWT-1KL111:53:D176AACXX:1:1106:4169:92324 419 chr1 11685 3 75M = 11790 180 GATTCTTATTAGTGATTTGGGCTGGGGCCTGGCCATGTGTATTTTTTAAATTTCCACTGATGATTTTGCTGCAT **R**? @DBDDDHHBDAFGIIJJIEHJJJE<GGHGGBH@?D?DGHFGEGD@AHGIHGGGDGIIICEH:?EHFBDF@### AS: i:0 XN:i:0 XM: i:0 X0:i:0 XG:i:0 NM:i:0 MD:7:75 YT:7:UU NH:i:2 CC:7:chr15 CP:i:102519411 HT:i:0 HWI-1KL111:53:D176AACXX:1:1303:20062:7739 419 chr1 11708 3 75M = 11771 138 GGGGCCTGGCCATGTGTATTTTTTTAAATTTCCACTGATGATTTTGCTGCATGGCCGGTGTTGAGAATGACTGCN @<BBFFFDHFHB;ECGDGGHGIGGGEFBFHIGGGHII<FH@FEGHGBGGDFFHJICHGHHEFB@C;A@########</pre> AS: i: -1 XN: i:0 XM: i:1 X0:i:0 XG:i:0 NM:i:1 MD:7:74G0 YT:7:UU NH:i:2 CC:7:chr15 CP:i:102519388 HT:i:0 HWI - 1KL 111:53: D176AACXX: 1:2105:1895:21247 339 75M 11650 -143 chr1 11718 3 = CATGTGTATTTTTTAAATTTCCACTGATGATTTTGCTGCATGGCCGGTGTTGAGAATGACTGCGCAAATTTGCC 9<0?:=EHHDHCEE@CIGIHCCDHFB2HDGAFDD<G@F?FCC@DHAFHBGGAC8:AFHFDDDDD@@< AS:i:0 XN:i:0 XM:i:0 X0:i:0 XG:i:0 NM:i:0 MD:Z:75 YT:Z:UU NH:i:2 CC:Z:chr15 CP:i:102519378 HI:i:0

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- Expression is usually measured in Reads Per Kilobase of transcript per Million mapped reads (RPKM).



A pipeline to study gene expression and isoform expression

One of the most commonly used workflows is to map reads with a tool like *Tophat* and then use a tool like *HTSeq* to count the number of reads overlapping a gene.



Integrated Genome Viewer

Some reads fall within junctions. First half falls on one exon and the second half on the other exon.



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Overview of RNA-SEQ analysis workflow



Common goals of RNA-Seq analysis

• differential expression of genes,

- differential expression of genes,
- differently spliced transcripts,

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- non-coding RNAs,

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- differently spliced transcripts,
- non-coding RNAs,
- post-transcriptional mutations,
- gene fusions.

• building fast and reliable read aligners,

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- transcriptome reconstruction (genome-guided, genome-independent),

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- transcriptome reconstruction (genome-guided, genome-independent),
- expression quantification (gene or isoform quantification, differential expression).

Questions of Interest

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RNA-SEQ experiments allow us to answer several questions about sequenced transcripts.

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- Expression levels (from counts over exons and junctions)
- Structure (e.g.: isoforms from the alignments)
- Variants (e.g.: Single-Nucleotide Polymorphism, indels)

• Characterize the full genome of an organism.

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- Discover unknown genes (possibly also non-coding ones).

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- Characterize transcript isoforms.
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- Monitor transcriptome changes across tissues or in response to environmental changes.
- Identify significant gene expression changes across different environmental conditions.
- Study what is encoded in a genome and how is it processed.

• Annotated regions stored in classical databases (e.g.: RefSeq, UCSC, Ensembl).

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- Boundary annotations (e.g.: exon boundaries, splicing sites, UTR's).
- Novel transcribed regions (e.g.: new exons, new isoforms).

Transcriptome Analysis

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Therefore:

- How can we take into account all this information?
- How can we can handle all this information?

Open research topics in Statistics for RNA-SEQ

Statistical analysis usually makes use of Poisson and Negative Binomial distributions.

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Novel Probabilistic models for RNA-SEQ are still needed.

De novo transcriptome assembly

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- *De novo transcriptome assembly* is the method of creating a transcriptome without the aid of a reference genome.

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Alignment strategies of RNA-SEQ reads to the genome



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Transcriptome reconstruction methods



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RNA-SEQ

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Gene espression quantification with RNA-SEQ



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Graph theory to locally assembly genes/transcripts



Figure:

Two strategies for genome assembly: from Hamiltonian cycles to Eulerian cycles



Differential Expression



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